Influence of Triple-Lumen Central Venous Catheters Coated With Chlorhexidine and Silver Sulfadiazine on the Incidence of Catheter-Related Bacteremia

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Objective: To evaluate the efficacy of triple-lumen central venous catheters coated with a combination product of chlorhexidine and silver sulfadiazine (CSS) in reducing the incidence of local catheter infection and catheter-related bacteremia.

Design: Randomized, controlled trial.

Setting: The surgical intensive care units in a university hospital.

Patients: All patients who needed central venous catheterization were randomized to receive either an uncoated triple-lumen catheter (n=157) or a catheter coated with CSS (n=151).

Main Outcome Measure: Catheters were removed when no longer needed or suspected as a cause of infection. The tip and a 5-cm segment of the intradermal portion of the catheter were cultured semiquantitatively. Blood cultures were obtained when clinically indicated. The remaining segment of catheters coated with CSS were cut and incubated on an agar plate with strains of Staphylococcus aureus and Enterococcus. Zone of inhibition was determined 24 hours later. Data were analyzed by survival and logistic multivariate regression methods.

Results: Catheters coated with CSS were effective in reducing the rate of significant bacterial growth on either the tip or intradermal segment (40%) compared with control catheters (52%; P=.04). However, there was no difference in the incidence of catheter-related bacteremia (3.8% [uncoated] vs 3.3% [coated]; P=.81). In vitro activity of catheters with CSS against S. aureus was evident up to 25 days but activity against Enterococcus dissipated more quickly over time and was absent by day 4. The most common colonizing organisms were coagulase-negative staphylococcus and enterococcus. Variables that were associated with a significant amount of growth on the tip or intradermal segment were a duration of catheterization of longer than 7 days, jugular insertion site, and the absence of a CSS coating. The use of a guidewire when the catheter was removed was associated with a lower risk of significant bacterial growth.

Conclusions: The use of CSS reduces the incidence of significant bacterial growth on either the tip or intradermal segments of coated triple-lumen catheters but has no effect on the incidence of catheter-related bacteremia. In this patient population, catheters coated with CSS provide no additional benefit over uncoated catheters.

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Despite improvement in central venous catheter (CVC) design and insertion techniques, catheter-related infections continue to be a significant problem in the intensive care unit.1-3 One popular strategy to reduce these infections is scheduled CVC change over a guidewire; however, several studies4-5 have indicated that this approach does not reduce the risk of infection. A collagen cuff impregnated with silver that fits around the CVC and resides in a subcutaneous pocket was shown to reduce the density of growth of bacteria on the catheter in some but not all studies.6-9 None of these studies individually demonstrated a statistically significant reduction in the incidence of catheter-related bacteremia (CRB); however, pooled data from 2 studies6,7 suggest that the use of the cuff resulted in a significant decrease in the rate of CRB.7 This product has not gained widespread popularity possibly because the use of the cuff can be cumbersome and a substantial portion of these cuffs extrude from their pockets over time. Antibiotic-bonded CVCs have been used in several animal and clinical studies,10-13 but concerns about the emergence of resistant organisms and the time needed to bond the catheter with antibiotics at the bedside has limited the use of this approach. The CVC coated with the antiseptic combination product of chlorhexidine and silver sulfadiazine (CSS)
MATERIALS AND METHODS

This study was reviewed and approved by the University of Massachusetts Medical Center Committee for the Protection of Human Subjects in Research in Worcester. Since both products (Arrow-Howes multilumen catheter and Arrowgard multilumen catheter, Arrow International Inc, Reading, Pa) are commercially available and no deviation from routine clinical protocols was made, informed consent was waived. All patients who were admitted to the surgical intensive care units at the University of Massachusetts Medical Center from March 1994 through June 1995 and who needed a CVC were eligible for the study. Randomization was based on the last digit of the medical record number: patients with an odd number received a standard uncoated CVC.

A total of 380 patients were planned to be enrolled into the study. A power analysis indicated that to observe a reduction in significant catheter colonization from 30% (our endemic rate) to 15% would require approximately 140 patients in each group. An additional 50 patients per group were added to allow for those who discontinued the study. Such a sample size, however, would be unlikely to detect changes in CRB rates (power approximately 10%).

CATHETER CARE

Catheters were inserted in external or internal jugular, subclavian, or femoral veins. Use of guidewires was allowed. House staff inserted all the catheters; use of masks, caps, gowns, gloves, and drapes was required.19,20 Prior to insertion, the skin was cleansed with a 10% povidone-iodine solution. A transparent dressing (Polyskin 2, Kendall Corporation, Mansfield, Mass) was used to cover the catheter at the insertion site; no antibiotic ointment was used. Dressings were changed every 48 hours or sooner at the discretion of the nurse caring for the patient and the site was cleaned with a solution of alcohol and acetone.

CULTURES

When the catheters were changed or removed, portions of the tip and intradermal segment were sent for semiquantitative culture by a technician in the hospital microbiology laboratory. In the laboratory, the segments were cut to approximately 5 cm and rolled 4 times on a 5% sheep blood agar plate. The plates were incubated in 5% carbon dioxide at 35°C to 37°C. Colony-forming units (CFUs) were counted at 48 hours.21 If there was a suspicion of bacteremia, peripheral venous blood cultures were obtained and sent to the hospital microbiology laboratory. For the coated CVC, the remaining portions were sent to a research microbiology laboratory, where they were flushed with nonbacteriostatic saline solution, cut, measured, weighed, and placed on Mueller-Hinton agar plates inoculated with a suspension (0.5 McFarland standard or approximately 1.5×10^9 CFUs/mL) of Staphylococcus aureus, which has been used in an animal model of catheter infection22 and of Enterococcus (a blood isolate from a patient). The plates were incubated for 24 hours at 35°C in ambient air. Zone of inhibition of S aureus and Enterococcus growth was measured. Significant growth was defined as the tip or intradermal segment growing more than 14 CFUs.23 Catheter-related bacteremia was defined as the isolation of the same bacteria from either the tip or intradermal segment (in significant quantities [>4 CFUs]) and the blood. The antibiogram was used to determine if bacteria were the same isolates.

DATA ANALYSIS

Other data that were collected included duration of catheterization, use of guidewires, site of catheterization, use of other catheters, use of total parenteral nutrition, and use of antibiotics. Data were analyzed using life-table analysis, x^2 test, Student t test, and multivariate logistic regression analysis (Statistica, Statsoft, Tulsa, Okla) where appropriate. Differences in the life-table analyses were considered significant if P<.05. Curve fitting was accomplished using a graphics software program (Fig P, Biosoft, Ferguson, Mo). In the multivariate logistic regression analysis, univariate analyses were performed first with variables that could have an impact on catheter colonization or infection. To avoid rejecting variables that could be important in catheter infection, the P value was set at .25.24 Variables that were found to be significant in the univariate analyses were included in the multivariate analysis where the P value for each variable was set at .05.

RESULTS

Three hundred sixty-five catheters were inserted in 251 patients. The characteristics of the patient population and catheters are presented in Table 1. There are no significant differences between the 2 groups. Fifty-seven catheters were removed or exchanged with other catheters and not cultured because the patient was transferred to a rehabilitation facility with the catheter in place or the patient had been transferred to a different area of the hospital (eg, operating room or ward) and the intensive care unit team was not notified that the catheter was being removed. The mean (±SEM) duration of catheterization was 9.0±0.6 days (range, 1-44 days) for the uncoated catheters vs 8.5±0.6 days (range, 1-65 days) for the coated group (P=.55). There was no significant difference in the sites of insertion between the 2 groups (54% subclavian sites for uncoated catheters vs 51% for coated catheters) (P>.05).

Fifty-two percent of either the tips or intradermal segments cultured from uncoated CVCs exhibited growth of more than 14 CFUs compared with 40% of either the tips or intradermal segments from CVCs with CSS, re-
Table 1. Characteristics of Patients and Catheter Use

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>Uncoated (n=157)</th>
<th>Coated (n=151)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mean (±SD) age, y</td>
<td>56.2±1.5</td>
<td>56.7±1.7</td>
</tr>
<tr>
<td>Sex (M:F), %</td>
<td>54:46</td>
<td>65:35</td>
</tr>
<tr>
<td>Admitting service, %</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Trauma</td>
<td>33</td>
<td>32</td>
</tr>
<tr>
<td>Vascular surgery</td>
<td>18</td>
<td>16</td>
</tr>
<tr>
<td>General surgery</td>
<td>28</td>
<td>32</td>
</tr>
<tr>
<td>Neurosurgery</td>
<td>13</td>
<td>9</td>
</tr>
<tr>
<td>Other</td>
<td>8</td>
<td>11</td>
</tr>
<tr>
<td>Arterial catheter present, %</td>
<td>85</td>
<td>89</td>
</tr>
<tr>
<td>Use of guidewire for exchanging</td>
<td>46:54</td>
<td>49:51</td>
</tr>
<tr>
<td>catheters (yes:no), %</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mean (±SD) white blood cell</td>
<td>14.5±1.7</td>
<td>13.3±0.5</td>
</tr>
<tr>
<td>count on catheter removal, ×10⁹/L</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Temperature on removal, °C</td>
<td>37.9±0.1</td>
<td>37.8±0.1</td>
</tr>
<tr>
<td>Use of total parenteral nutrition</td>
<td>18</td>
<td>23</td>
</tr>
<tr>
<td>Use of antibiotic(s), %</td>
<td>91</td>
<td>97</td>
</tr>
</tbody>
</table>

Figure 1. Cumulative percentage of catheter segments (either tips or intradermal segments) with less than 15 colony-forming units of bacterial growth as a function of duration of catheterization. The 2 curves are not significantly different from each other (P=.13).

Figure 2. Cumulative percentage of cases without catheter-related bacteremia. There are no significant differences between the 2 curves.

One of the major findings of this study is that CVCs coated with CSS are effective in reducing the risk of developing significant bacterial growth on the catheter. However, this efficacy does not become apparent until the catheter has been in the patient for approximately 7 days or longer (Figure 1) and does not translate into a reduced incidence of catheter-related bloodstream infection. Since antimicrobial activity drops off significantly within the first 5 to 7 days (Figure 3, top), it is not immediately clear why
Infection the longer a catheter remains in place.\(^24,25\)

Importance in the pathogenesis of catheter-related infection. Luminal colonization assumes much greater importance in the pathogenesis of catheter-related infection. Catheter samples only from the external surface of the catheter. The patient realize a benefit from a CSS catheter. The duration of catheterization is beyond 7 to 10 days will result in a clinically beneficial effect. This ability of CSS to prevent the formation of biofilm may not result in a clinically beneficial effect. The use of quantitative culturing methods that detect both external and luminal bacterial growth should be the same. As a result, it is unlikely that the use of quantitative culturing methods that detect both external and luminal bacterial growth would have greatly altered our findings.

Our findings support some of the results of previous studies published in abstract form and one small, recently published study. Van Heerden et al\(^26\) showed that triple-lumen catheters coated with CSS reduced the incidence of significant bacterial growth on the tip by 63%. Maki et al\(^19\) demonstrated that catheters coated with CSS were effective in reducing significant bacterial growth on the catheter and the incidence of CRB compared with uncoated catheters. Similar data have been presented by other groups.\(^15,16,27\)

Irrespective of the type of catheter, the incidence of significant bacterial growth is higher in this study compared with others.\(^3,6,12,14,15,15,27-29\)

The reason for this higher rate is not clear but may be explained in part by the fact that we combined the colonization rates for both the tips and intradermal segments and the duration of catheterization was longer in this study.\(^5,6,12\)

Maximal sterile barrier precautions (ie, caps, masks, sterile gowns, gloves, and large sterile drapes) have been shown to reduce the incidence of contamination at the time of insertion and prevent subsequent catheter infection.\(^19,20\)

Although masks, caps, sterile gowns, gloves, and the sterile drape supplied in the catheter kit were used, sterile drapes that cover the entire patient and bed, as described by Raad et al,\(^20\) were not used. This may explain our higher rate of significant bacterial growth. We used transparent occlusive dressings for catheter care. A number of recent studies, including a meta-analysis,\(^30\) suggest that the use of dressings of this type predispose to enhanced bacterial growth on the skin, which translates into a higher risk of local catheter infection and CRB. However, the dressings were changed every 48 hours or sooner. The risk of local infection does not appear to be increased if these types of dressings are changed that frequently.

Use of guidewires to exchange catheters was allowed in this study. Theoretically, such a practice might predispose to a higher colonization rate. The univariate logistic regression analysis failed to identify the use of guidewire exchange during catheter insertion as a contributing factor to significant bacterial growth, a finding supported by some\(^5,31,32\) but not all investigations.\(^3,33-35\)

### Table 2. Number and Types of Isolates From Cultured Catheter Tips or Intradermal Segments\(^*\)

<table>
<thead>
<tr>
<th>Organism</th>
<th>Uncoated Coated</th>
<th>Organism</th>
<th>Uncoated Coated</th>
</tr>
</thead>
<tbody>
<tr>
<td>Staphylococcus aureus</td>
<td>2</td>
<td>Pseudomonas aeruginosa</td>
<td>3</td>
</tr>
<tr>
<td>Coagulase-negative staphylococcus</td>
<td>103</td>
<td>Pseudomonas spp</td>
<td>1</td>
</tr>
<tr>
<td>Staphylococcus (unspecified)</td>
<td>1</td>
<td>Xanthomonas maltophilia</td>
<td>1</td>
</tr>
<tr>
<td>Enterococcus</td>
<td>21</td>
<td>Acinetobacter spp</td>
<td>7</td>
</tr>
<tr>
<td>Streptococcus spp</td>
<td>1</td>
<td>Diphtheroids</td>
<td>33</td>
</tr>
<tr>
<td>Escherichia coli</td>
<td>5</td>
<td>Bacillus spp</td>
<td>4</td>
</tr>
<tr>
<td>Enterobacter aerogenes</td>
<td>4</td>
<td>Candida albicans</td>
<td>3</td>
</tr>
<tr>
<td>Klebsiella pneumoniae</td>
<td>3</td>
<td>Candida parapsilosis</td>
<td>1</td>
</tr>
<tr>
<td>Serratia spp</td>
<td>3</td>
<td>Yeast</td>
<td>2</td>
</tr>
<tr>
<td>Proteus spp</td>
<td>3</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*Ellipses indicate not applicable.

### Table 3. Microbiology of Cases of Catheter-Related Bacteremia

<table>
<thead>
<tr>
<th>Bacteria Isolates</th>
<th>No. of Cases</th>
</tr>
</thead>
<tbody>
<tr>
<td>Coagulase-negative staphylococcus</td>
<td>1</td>
</tr>
<tr>
<td>Enterococcus</td>
<td>2</td>
</tr>
<tr>
<td>Enterobacter cloacae</td>
<td>1</td>
</tr>
<tr>
<td>Candida albicans</td>
<td>1</td>
</tr>
<tr>
<td>Staphylococcus aureus</td>
<td>1</td>
</tr>
<tr>
<td>Coagulase-negative staphylococcus</td>
<td>2</td>
</tr>
<tr>
<td>Enterococcus</td>
<td>3</td>
</tr>
<tr>
<td>Candida parapsilosis</td>
<td>1</td>
</tr>
</tbody>
</table>

*One case of catheter-related bacteremia was polymicrobial: coagulase-negative staphylococcus and Enterococcus.
However, the use of guidewire exchange when removing the catheter was associated with a lower risk of significant colonization. This finding is due to several reasons. The duration of catheterization when a guidewire was used to remove a catheter was shorter than when a guidewire was not used (mean ± SEM, 6.49 ± 0.74 days vs 9.20 ± 0.48 days; \( P = .01 \)). One would expect a lower incidence of significant catheter colonization if the duration of catheterization was shorter. However, this cannot be the only explanation since duration of catheterization and the use of a guidewire on catheter removal were important variables in the multivariate analysis. An examination of the multivariate analysis indicated that the use of the guidewire on catheter removal had only a moderate effect on duration of catheterization as a confounding variable. Another explanation for the lower risk is related to the method of catheter removal over a guidewire. In this situation, the insertion site and the catheter that was to be removed were prepared with copious amounts of 10% povidone-iodine solution. Any excess povidone-iodine solution could coat the catheter as it was removed, thereby reducing bacterial burden.

Several other variables were associated with the development of significant bacterial growth on the catheter tip or intradermal segment, including a jugular site of catheterization and absence of a CSS catheter. In addition, there was a trend suggesting that duration of catheterization of longer than 7 days was associated with a higher risk of significant growth. Other investigations have corroborated these observations. At least 2 studies have shown that jugular venous catheterization is associated with a higher risk of significant colonization. The reason for the higher rate of colonization for jugular venous catheterization remains obscure but may be related to difficulties in dressing the catheter, proximity of the insertion site to the mouth, and differences in the density of local skin flora. The lower risk of infection in the subclavian site must be tempered with the small but increased risk of a pneumothorax when that approach is used. A number of studies have demonstrated the increasing risk of infection the longer a catheter is left in place. Routine catheter changes do not appear to be efficacious in reducing the risk of catheter infection and recent guidelines from the Centers for Disease Control and Prevention, Atlanta, Ga, recommend against such a practice. Contrary to catheter colonization, no variable was significantly associated with the presence of a CRB, including the presence of a CSS catheter.

### Table 4. Univariate and Multivariate Analyses of Catheter Tip and Intradermal Segment Cultures and CRB

<table>
<thead>
<tr>
<th>Variable</th>
<th>Bacterial Growth &gt;14 CFUs</th>
<th>CRB</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Odds Ratio</td>
<td>95% Confidence Interval</td>
</tr>
<tr>
<td>Univariate Analysis</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Jugular site of insertion</td>
<td>3.46</td>
<td>2.16-5.53</td>
</tr>
<tr>
<td>Absence of a CSS coating</td>
<td>1.57</td>
<td>1.0-2.45</td>
</tr>
<tr>
<td>Catheterization &gt;7 d</td>
<td>1.43</td>
<td>0.92-2.27</td>
</tr>
<tr>
<td>Use of guidewire on removal of catheter</td>
<td>0.33</td>
<td>0.17-0.63</td>
</tr>
<tr>
<td>Multivariate Analysis</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Jugular site of insertion</td>
<td>3.63</td>
<td>2.23-5.93</td>
</tr>
<tr>
<td>Absence of a CSS coating</td>
<td>1.89</td>
<td>1.03-2.78</td>
</tr>
<tr>
<td>Catheterization &gt;7 d</td>
<td>1.54</td>
<td>0.93-2.5</td>
</tr>
<tr>
<td>Use of guidewire for catheter removal</td>
<td>0.36</td>
<td>0.18-0.73</td>
</tr>
</tbody>
</table>

*CRB indicates catheter-related bacteremia; CFU, colony-forming unit; CSS, combination of chlorhexidine and silver sulfadiazine; and ellipses, not applicable.

### Figure 3. Zone of inhibition of catheters coated with a combination product of chlorhexidine and silver sulfadiazine on Staphylococcus aureus (top) and Enterococcus (bottom) as a function of duration of catheterization. The diameter of the catheter is 2.5 mm; therefore, a zone of inhibition of 2.5 mm (dashed line) represents no inhibition of growth. Each dot represents 1 catheter segment (only 1 segment from each patient was used).
Although the incidence of significant bacterial growth on either the catheter tips or intradermal segments was high, the incidence of CRB was less than 5% in both groups and is comparable with previously published reports. More than 300 catheters were cultured, yet we were unable to detect any difference in the rate of CRB between the 2 groups. It is likely that the power of this study was insufficient to determine a difference between coated and uncoated catheters in the prevention of CRB. Assuming an α error of .05 and an α error of .20, for a 50% reduction in the rate of CRB (4%-2%), more than 1300 patients would be needed in each group. Such a study, if not impossible, would require substantial resources.

The CSS coating was generally effective in vitro against the test strain of S aureus. However, the antimicrobial activity against Enterococcus was much less robust and dissipated more quickly as a function of duration of catheterization. This result probably explains why there was no difference in the number of Enterococcus isolates for the CSS catheter compared with control catheters. The organism responsible for the largest number of catheters with significant colonization was coagulase-negative staphylococcus, a finding observed in other studies. Recently published data show that unused CSS catheters will inhibit the growth of Staphylococcus epidermidis in vitro. Although we did not test in vitro activity of CSS against these species as a function of duration of catheterization, the absolute number of isolates of coagulase-negative staphylococcus isolated from CSS catheters was reduced by approximately 30% compared with the uncoated catheters. The majority of CRB that occurred in patients irrespective of the type of catheter used was due to either coagulase-negative staphylococcus or Enterococcus, an indication that these bacteria will continue to be an important cause of nosocomial bacteremias even if CSS catheters are used.

The method used to identify CRB in this study may have overestimated the rate of bacteremia. Recent data show that molecular typing of bacteria will result in a more accurate determination of CRB. In this study, the use of molecular typing could have resulted in lower rates of bacteremia, but it is unlikely that the difference between CRB rates would change. All 3 cases of coagulase-negative staphylococcus CRB were diagnosed on the basis of only 1 positive blood culture. Traditionally, 2 positive blood cultures are required before a CRB with this organism can be diagnosed. However, even if these cases of CRB are excluded, the differences in the rates of bacteremia remain insignificant. These findings are in agreement with 2 recent studies evaluating the efficacy of CVCs coated with CSS in patients receiving total parenteral nutrition.

In summary, we demonstrated that central venous catheterization with triple-lumen catheters coated with CSS were effective in reducing significant bacterial growth compared with uncoated catheters. There was no apparent effect on the incidence of CRB. Activity of the coating on these catheters against enterococci appears limited. In our patient population, the use of the CSS catheter does not appear to provide any clinically significant benefit over uncoated catheters.

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REFERENCES


