**Clostridium difficile**–Associated Diarrhea

**A Review**

Eleftherios Mylonakis, MD; Edward T. Ryan, MD; Stephen B. Calderwood, MD

*Clostridium difficile* causes 300,000 to 3,000,000 cases of diarrhea and colitis in the United States every year. Antibiotics most frequently associated with the infection are clindamycin, ampicillin, amoxicillin, and cephalosporins, but all antibiotics may predispose patients to *C difficile* infection. The clinical presentation varies from asymptomatic colonization to mild diarrhea to severe debilitating disease, with high fever, severe abdominal pain, paralytic ileus, colonic dilation (or megacolon), or even perforation. The most sensitive and specific test available for diagnosis of *C difficile* infection is a tissue culture assay for the cytotoxicity of toxin B. However, this test takes 1 to 3 days to complete and requires tissue culture facilities. Detection of *C difficile* toxin by means of enzyme-linked immunoassay is more rapid and inexpensive. A minority of patients may require more than 1 stool assay to detect toxin. Oral metronidazole or oral vancomycin hydrochloride for 10 to 14 days are equally effective at resolving clinical symptoms; oral metronidazole is preferred in most cases because of lowered cost and less selective pressure for vancomycin-resistant organisms. Approximately 15% of patients experience relapse after initial therapy and require retreatment, sometimes with an extended, tapering regimen. Immunity appears to be incomplete and predominantly mediated by serum IgG to toxin A. Measures for preventing the spread of the pathogen, appropriate diagnostic testing, and treatment may avert morbidity and mortality due to *C difficile*–associated diarrhea.

Arch Intern Med. 2001;161:525-533

*Clostridium difficile*, a Gram-positive, spore-forming anaerobic bacillus, was first described in 1935,1 but it was not associated with antibiotic-related diarrhea until the late 1970s.2-4 *Clostridium difficile* can lead to severe complications and currently is the most common cause of nosocomial diarrhea (often adding up to 2 weeks to the length of the hospitalization, at an additional cost of $6000-$10,000 per case).5-10 This report focuses on new information on epidemiological features, pathogenesis, immunological findings, clinical manifestations, diagnosis, and treatment of *C difficile* infection.

**EPIDEMIOLOGICAL FEATURES**

*Clostridium difficile* is the cause of approximately 25% of all cases of antibiotic-associated diarrhea.11 In most cases of antibiotic-associated diarrhea in which *C difficile* is not detected, no etiologic agent is identified, and diarrhea is usually mild and not accompanied by abdominal pain.12 Symptoms in such cases usually respond to discontinuation of treatment with the offending antibiotic.

Most cases of *C difficile*–associated disease occur in hospitals or long-term care facilities (rate of 25-60 per 100,000 occupied bed-days), causing more than 300,000 cases per year in the United States alone.12-14 The incidence of this infection in the outpatient setting (7.7 cases per 100,000 person-years; approximately 20,000 cases per year in the United States) is lower, but not negligible.15 Overall, the risk for development of *C difficile*–associated diarrhea (CDAD) within 6 weeks of a course of antibiotics in the outpatient setting is low (6.7 cases per 100,000 exposures).15

Among hospitalized individuals, the risk for CDAD after clindamycin therapy has been estimated to range from 1 in 10...
with isolation of cases, reducing the environmental burden with appropriate use of disinfectants,20,29 and education of hospital personnel, can help control the outbreak.

PATHOGENESIS AND IMMUNOLOGICAL FEATURES

Infants and young children commonly harbor C difficile in fecal flora but have no symptoms related to production of toxin.20,30 For unclear reasons, the number of carriers quickly declines as children age, and toxigenic C difficile is isolated from stool specimens in only 0% to 3% of healthy adults.20 During hospitalization, however, colonization frequently occurs (21% of patients in one study, for example).13 Although asymptomatic carriers are an important hidden reservoir of C difficile, clinical symptoms develop in only about one third of colonized patients, and asymptomatic colonization with C difficile may be associated with a decreased risk for development of CDAD.31

Clostridium difficile forms spores that persist in the environment for years, and contamination by C difficile is common in hospitals and long-term care facilities, especially in rooms occupied by an infected individual.13,32-35 Patient-to-patient transmission of the organism occurs, and the organism can be cultured from many environmental surfaces in rooms of infected patients and on hands, clothing, and stethoscopes of health care workers. Hospital personnel may carry the bacteria from room to room and promote infection, but fecal carriage by staff is rare.13,32,34 Outbreaks can occur in hospitals, nursing homes, and other extended-care facilities.22

The first step in development of C difficile colonization is disruption of the normal flora of the colon, usually caused by antibiotics or, in unusual cases, by antineoplastic or immunosuppressive drugs.20,36 Colonization occurs by the fecal-oral route; ingested spores of C difficile survive the gastric acid barrier and germinate in the colon.20,22 Symptoms of CDAD may start on the first day of antibiotic therapy or 6 weeks or longer after antibiotic therapy is stopped.6

The following 2 factors recently have been shown to increase the probability of symptomatic disease in patients who acquire C difficile colonization in hospital: the severity of other illnesses, and reduced levels of serum IgG antibody to toxin A.37 These results suggest that pre-existing anti–toxin A antibody may ameliorate severity of disease and that immunization might be efficacious in preventing and controlling nosocomial CDAD.

Some strains of C difficile are nontoxigenic; these strains do not cause disease. Clinically significant strains produce 2 protein exotoxins, toxin A, a 308-kd protein, and toxin B, an approximately 270-kd protein; these toxins are largely responsible for disease manifestations. However, rare cases of CDAD caused by toxin A–negative, toxin B–positive strains have been reported.38,39 The genes encoding both toxins are located very near each other on a 19-kilobase-pair pathogenicity island in toxigenic strains.20 Toxin A binds to a specific receptor on the brush border of the intestinal epithelium, a glycoprotein with an α-linked galactose; the intestinal receptor in humans for toxin B is less well characterized.40 After binding to appropriate receptors, the toxins are internalized and act within the eukaryotic cell to modify proteins covalently in the Rho subfamily, a group of low-molecular-weight GTP-binding proteins involved in regulation of the F-actin cytoskeleton.40 Both toxins modify Rho proteins at a specific threonine residue by addition of a glucose molecule, leading to inactivation of the protein.40-45 This is followed by disaggregation of polymerized actin, opening of tight junctions between cells, cell rounding, and subsequent cell death. The toxins also induce the release by various cells of proinflammatory mediators and cytokines, as well as activation of the enteric nervous system, leading to neutrophil chemotaxis and fluid secretion.46 Toxin B is not enterotoxic in animals (as is toxin A), but it is a much more potent cytotoxin in tissue culture than toxin A, and full tissue damage requires the action of both toxins.22,47

Early histological changes due to C difficile toxins include patchy epithelial necrosis with an exudate of neutrophils and fibrin; the exudate

### Table 1. Antimicrobial and Antineoplastic Agents That Induce Clostridium difficile–Associated Diarrhea and Colitis*

<table>
<thead>
<tr>
<th>Common</th>
<th>Antimicrobial and Antineoplastic Agents</th>
</tr>
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<tbody>
<tr>
<td>Clindamycin</td>
<td>Ampicillin and amoxicillin</td>
</tr>
<tr>
<td>Cephalosporins</td>
<td></td>
</tr>
<tr>
<td>Less common</td>
<td>Chloramphenicol</td>
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<tr>
<td></td>
<td>Co-trimoxazole</td>
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<tr>
<td></td>
<td>Erythromycin and other macrolides</td>
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<tr>
<td>Penicillins (other than ampicillin or amoxicillin)</td>
<td></td>
</tr>
<tr>
<td>Tetracyclines</td>
<td>Trimethoprim sulfamethoxazole</td>
</tr>
<tr>
<td>Quinolones†</td>
<td></td>
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<tr>
<td>Rare or questionable</td>
<td>Bacitracin</td>
</tr>
<tr>
<td></td>
<td>Cisplatin</td>
</tr>
<tr>
<td></td>
<td>Doxorubicin hydrochloride</td>
</tr>
<tr>
<td></td>
<td>Fluorouracil</td>
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<tr>
<td></td>
<td>Methotrexate</td>
</tr>
<tr>
<td></td>
<td>Parenteral aminoglycosides</td>
</tr>
<tr>
<td></td>
<td>Parenteral metronidazole</td>
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<tr>
<td></td>
<td>Parenteral vancomycin</td>
</tr>
<tr>
<td></td>
<td>Rifampin</td>
</tr>
<tr>
<td></td>
<td>Sulphonamides</td>
</tr>
<tr>
<td></td>
<td>Tacrolimus (FK506)</td>
</tr>
<tr>
<td></td>
<td>Teicoplanin</td>
</tr>
</tbody>
</table>

*Data from Anand et al,6 Schwaber et al,19 Kelly and LaMont,6 Anand and Glatt,21 Kelly et al,22 Manabe et al,23 Barbur et al,24 McFarland et al,25 and Riley et al.26 Because the differences among the nonbase moieties of each antimicrobial agent have not been studied for the development of C difficile–associated diarrhea, nonbase moieties are not given.

†Data on the newer fluoroquinolones with better activity against Gram-positive bacteria are limited.27
may appear to “erupt” into the colonic lumen and, if prominent, such a histological feature has been termed a summit or volcano lesion. More prominent histological changes are characterized by diffuse epithelial necrosis and ulceration with development of an overlying pseudomembrane containing cellular debris, leukocytes, fibrin, and mucin. (Figure 1).

In animal models, antibodies directed against toxin A prevent toxin binding, neutralize secretory and inflammatory effects, and limit or prevent clinical disease. Antibodies against C difficile toxins are present in most adults and older children (>60%). The presence of such antibodies may be due to repetitive, transient presence of toxigenic C difficile organisms in the normal intestinal flora. Cellular immunity appears to be of lesser import, but has not been studied in detail.

**CLINICAL MANIFESTATIONS**

The clinical presentation of CDAD is variable and includes diarrhea, colitis without pseudomembranes, pseudomembranous colitis, and fulminant colitis (Table 2). Some individuals with toxigenic strains in stool remain totally asymptomatic. Mild to moderate CDAD is usually accompanied by lower abdominal cramping pain but no systemic symptoms or physical findings. Moderate or severe colitis usually presents with profuse diarrhea, abdominal distention with pain, and, in some cases, occult colonic bleeding. Also, systemic symptoms such as fever, nausea, anorexia, and malaise are usually present. A minority of patients have disease primarily in the cecum and right colon, presenting with marked leukocytosis and abdominal pain but little or no diarrhea.

Fulminant colitis develops in approximately 1% to 3% of patients, with ileus, toxic megacolon, perforation, and death. Clinicians should be aware that the development of these life-threatening complications may be accompanied by a decrease in diarrhea due to loss of colonic muscular tone and ileus.

Other complications of C difficile infection include hyperpyrexia, chronic diarrhea, and hypobulinemia with anasarca. Clostridium difficile infection may occasionally complicate idiopathic inflammatory bowel disease. Also, a reactive
arthritis occurring 1 to 4 weeks after C difficile colitis develops in some patients, resembling that seen after other enteric infections such as Yersinia, Shigella, and Salmonella disease. 99

**DIAGNOSIS**

Non-specific laboratory abnormalities in patients with CDAD include leukocytosis with a shift to the left and fecal leukocytes in about 50% to 60% of cases. The average peripheral white blood cell count is $12 \times 10^9/L$ to $20 \times 10^9/L$, but occasionally the peripheral count is higher and cases of leukemoid reaction have been described. 4,3,11,70

Even among patients with the most severe colitis, C difficile constitutes only a small part of the colonic flora, and its morphologic features are identical to that of other Clostridia species, making Gram staining of fecal specimens of no value in diagnosing CDAD. Clostridium difficile can be isolated by means of anaerobic culture of stool, but this test is seldom used for clinical diagnosis (it takes 2-3 days to complete and does not distinguish toxigenic from nontoxigenic strains; nontoxigenic strains are not associated with clinical illness).

The most sensitive and specific test available for diagnosis of C difficile infection is a tissue culture assay for the cytotoxicity of toxin B, using preincubation with neutralizing antibody to show the specificity of the cytotoxicity. 22,27 This test can detect as little as 10 pg of toxin in stool and has a high sensitivity (94%-100%) and specificity (99%). 50,22 However, the test takes 1 to 3 days to complete and requires tissue culture facilities.

More recently, enzyme-linked immunosorbent assays (ELISAs) have been developed to detect toxin A and/or B in stool. These assays detect 100 to 1000 pg of either toxin, and have a sensitivity of 71% to 94% and a specificity of 92% to 98%. 71,72 Because of the rapidity of testing and ease of performance, ELISAs for toxins A and B are now used most frequently by clinical laboratories for diagnosis of C difficile infection. Approximately 5% to 20% of patients may require more than 1 stool assay to detect toxin. 25 If CDAD is suspected, a single stool specimen should be sent. If the results are negative and diarrhea persists, 1 or 2 additional stool samples can be sent. 73,74 The cytotoxicity assay for toxin B will detect an additional 5% to 10% of cases missed by ELISA techniques, and it may be useful to perform this test if the ELISA results are negative but clinical suspicion is high (Table 3). 20

Diagnostic testing during or at the end of treatment or during follow-up is not needed, unless symptoms recur. 75 Transmission of infection to other patients is associated with ongoing diarrhea and not with the presence of the toxin in stool. Enteric precautions can be removed when diarrhea subsides, without the need for repeated diagnostic testing. 73

Studies have also evaluated other laboratory methods for the diagnosis of CDAD. The polymerase chain reaction is extremely sensitive but requires technical expertise and is unable to distinguish between asymptomatic carriage and symptomatic infection. 75 A dot-immunobinding assay yielded fair initial results several years ago, but this test has had very limited clinical experience since. 76 Counterimmunoelectrophoresis for detection of C difficile antigens has demonstrated poor positive predictive value, and a latex agglutination test is less sensitive than other available tests; newer forms of ELISAs are under evaluation. 77,78

Radiographic imaging studies can be used to assist in making a diagnosis of CDAD. Abdominal imaging studies may reveal paralytic ileus and a dilated colon (Figure 2). Diffusely thickened or edematous colonic mucosa may sometimes be seen by means of abdominal computerized tomographic scans (Figure 3); such thickening can present as “thumbprinting” on abdominal plain films.

Endoscopy for CDAD is reserved for special situations, such as when other diseases need to be ruled out, rapid diagnosis is necessary, or a stool sample cannot be obtained because ileus develops. 74 The results of sigmoidoscopy may be normal in patients with mild disease. Endoscopy in more severe colitis without pseudomembrane formation usually reveals diffuse or patchy, non-specific colitis. 22 In more dramatic cases, endoscopy may demonstrate characteristic, adherent yellow plaques that vary in diameter from 2 to 10 mm and in some cases, coalesce to cover large areas of the mucosa (Figure 4). In most cases, the rectum and sigmoid are involved, but 10% of episodes of colitis involve only the right colon and may not be detected by flexible sigmoidoscopy. 79 Due to the increased risk for intestinal perforation, endoscopy should be used sparingly as a diagnostic tool in patients with suspected CDAD.

**TREATMENT**

Practical guidelines for the treatment of CDAD are summarized in Table 4. The inciting antibiotic therapy should be discontinued if

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*Table 3. Diagnostic Tests for Detecting Clostridium difficile*

<table>
<thead>
<tr>
<th>Test</th>
<th>Sensitivity</th>
<th>Specificity</th>
<th>Ease of performance</th>
<th>Time required for results</th>
</tr>
</thead>
<tbody>
<tr>
<td>Enzyme-Linked Immunosorbent Assay</td>
<td>++</td>
<td>++</td>
<td>+++/++++</td>
<td>+++</td>
</tr>
<tr>
<td>Cytotoxin Assay</td>
<td>+++</td>
<td>+/++</td>
<td>+++/++++</td>
<td>+++</td>
</tr>
<tr>
<td>Latex Particle Agglutination</td>
<td>++</td>
<td>+++</td>
<td>+++/++++</td>
<td>+++</td>
</tr>
<tr>
<td>Culture</td>
<td>+++</td>
<td>++</td>
<td>+++/++++</td>
<td>+++</td>
</tr>
<tr>
<td>Polymerase Chain Reaction†</td>
<td>+++</td>
<td>++</td>
<td>+++/++++</td>
<td>+++</td>
</tr>
</tbody>
</table>

*†Initial results fair but still not widely used.
‡Allows strain typing in epidemics.

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possible, and supportive therapy should be given with fluids and electrolytes as needed. Education of hospital personnel and infection control issues are of paramount importance in the management of CDAD. Education should reinforce the importance of handwashing between patients and the need for glove use for the handling of bodily substances of all patients. Hospital personnel should thoroughly disinfect objects contaminated with *C. difficile* and should be familiar with the disease and its epidemiological features. Enteric isolation precautions are recommended for patients with CDAD, and patients should be moved to a private room if possible.

Antiperistaltic and opiate drugs should be avoided in patients with CDAD, because they mask symptoms and may worsen the course of the disease. Diarrhea will resolve without specific antimicrobial therapy in up to one fourth of patients.

Antimicrobial therapy for CDAD is indicated for patients with moderate or severe disease or significant underlying conditions. There is no evidence that treatment of asymptomatic carriers of *C. difficile* provides any clinical benefit, and such therapy has been associated with a prolongation of the carrier state. In the appropriate clinical setting, antimicrobial therapy should be instituted even before the laboratory results for *C. difficile* are available.

Oral metronidazole and vancomycin hydrochloride are the antibiotics most commonly used, whereas bacitracin, methylene disalicylate, teicoplanin, and fusidic acid also have some clinical efficacy. *Clostridium difficile* is uniformly susceptible to vancomycin, but occasional strains have been reported that are resistant to metronidazole. Two studies have directly compared initial therapy consisting of metronidazole (in an oral dose of 500 mg 3 times per day or 250 mg 4 times per day [qid]) with oral vancomycin hydrochloride (500 mg qid), each given for 10 days, and have shown that these therapies are equally efficacious in resolving symptoms as well as in the subsequent risk for relapse. Another study examined different doses of vancomycin hydrochloride and showed that an oral dose of 125 mg qid was equivalent to one of 500 mg qid for 10 days for symptomatic response and risk for subsequent relapse. In many series of patients, therapy with metronidazole or vancomycin was effective in
Table 4. General Guidelines for the Management of *Clostridium difficile*-Associated Diarrhea*

<table>
<thead>
<tr>
<th>Week</th>
<th>Vancomycin Hydrochloride Dose</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>125 mg qid</td>
</tr>
<tr>
<td>2</td>
<td>125 mg twice a day</td>
</tr>
<tr>
<td>3</td>
<td>125 mg every day</td>
</tr>
<tr>
<td>4</td>
<td>125 mg every other day</td>
</tr>
<tr>
<td>5 and 6</td>
<td>125 mg every 3 days</td>
</tr>
</tbody>
</table>

*Adapted from Johnson and Gerding* and Fekety.*

Isolate the patient.
2. Educate personnel to use gloves when in contact with patient and for the handling of bodily substances.
3. If possible, discontinue inciting antibiotic therapy and avoid antiperistaltic and opiate drugs.
4. Confirm the diagnosis with a test for *C difficile* toxin. If the results of the first specimen are negative and diarrhea persists, 1 or 2 additional stool samples should be sent.
5. If clinically indicated (moderate or severe diarrhea, systemic symptoms, significant leukocytosis, etc), consider antimicrobial treatment against *C difficile*. If the clinical suspicion is high and the patient is severely ill, empiric antimicrobial treatment may be started awaiting laboratory confirmation.
6. Oral metronidazole (250 mg 4 times per day or 500 mg 3 times per day) for 10-14 d is usually adequate.
7. Oral vancomycin hydrochloride (125 mg 4 times per day) for 10-14 d is indicated for those who cannot tolerate oral metronidazole, those in whom metronidazole therapy fails, pregnant patients, and, perhaps, severely ill patients.
8. The first relapse/recurrence of *C difficile* colitis can be treated with another 10- to 14-d course of oral metronidazole or vancomycin.
9. Therapy of patients with multiple relapses of *C difficile* colitis has not been examined by randomized, prospective, controlled clinical trials. A tapering course of metronidazole or vancomycin for 4-6 wk has been used.

Resolving symptoms in more than 95% of patients, although 10% to 20% of patients subsequently experienced relapse.*

Because of lower cost and avoidance of selective pressure for vancomycin-resistant organisms such as vancomycin-resistant enterococci, initial therapy with oral metronidazole (either 250 mg qid or 500 mg 3 times per day) is currently the preferred initial therapy for *C difficile* colitis. However, some authorities prefer initial therapy with vancomycin in the most severely ill patients, or in women who are pregnant. The duration of initial therapy should be 10 to 14 days (or therapy may be continued until 1 week after completion of the inciting antibiotic therapy, if it cannot be stopped earlier).

Metronidazole is well absorbed from the small intestine and produces low or undetectable levels in stool as *C difficile* diarrhea subsides, whereas oral vancomycin is not absorbed and produces high fecal levels even in the absence of diarrhea. In the presence of severe ileus or toxic megacolon, intravenous metronidazole hydrochloride (given as 500 mg every 8 hours) produces fecal concentrations above the inhibitory concentration for *C difficile* and may be used for initial therapy. Patients receiving metronidazole should avoid alcohol, because it may result in disulfiramlike reactions.

Metronidazole resistance in *C difficile* has been reported, but it is probably rare.

Oral bacitracin methylene disalicylate (20000 to 25000 U qid) has been shown to be as effective as vancomycin in ameliorating clinical symptoms of *C difficile* disease, but it was less effective in eradicating the organism or toxin from stool; patients treated with bacitracin also had a higher risk for subsequent relapse. Bacitracin should be reserved for unusual situations, in which metronidazole and vancomycin cannot be used.

In critically ill patients who are unable to take oral antimicrobials, treatment is empirical and may include intravenous metronidazole or administration of vancomycin by means of rectal enema or through long catheters in the small intestine. Intravenous treatment can be used with metronidazole but not vancomycin. Occasionally, surgery (usually subtotal colectomy) is required.

Despite the high initial response rate to therapy, recurrence of symptoms develops in 10% to 20% of patients, usually within the first 2 weeks after initial treatment is discontinued. Two studies attempted to identify possible risk factors for recurrent CDAD. High-risk groups included patients with chronic renal failure, those with multiple previous episodes of CDAD, individuals who had to continue other antibiotic therapy, patients with community-acquired CDAD, those with high white blood cell counts (≥15 × 10⁹/L), and those infected with certain strains of *C difficile*. About 50% of all recurrences of symptoms are actually due to reinfection and not to relapse of the primary infection.

The first relapse or recurrence of *C difficile* colitis can be treated with another 10- to 14-day course of oral metronidazole or vancomycin, either of which produces a response rate of approximately 95%; mild disease can be managed without further antibiotic treatment.

A smaller number of patients have multiple relapses. One study showed a relapse rate of 24% after an initial episode of *C difficile* colitis, but a relapse rate of 65% after treatment of recurrent disease. Antimicrobial resistance as the mechanism for relapse has not been documented, and one hypothesis is that spores of *C difficile* persist in the colon or the environment of the patient and that subsequent germination in the intestine produces vegetative forms and clinical illness. Therapy for multiple relapses of *C difficile* colitis has not been examined by randomized, prospective, controlled clinical trials and the best therapeutic approach is currently uncertain. A tapering course of metronidazole or vancomycin during a 4- to 6-week period has been used. A 6-week tapering course of vancomycin, as seen in the following tabulation, may allow spores to germinate during days between vancomycin therapy, with killing of vegetative forms on reexposure.
cholesteryramine and vancomycin (as well as other drugs) by at least 3 hours. Oral vancomycin hydrochloride, 125 mg qid, and oral rifampin, 600 mg twice daily for 7 days, may have some increased efficacy compared with vancomycin alone. Relapsing C difficile disease has also been treated with a combination of Saccharomyces boulardii, beginning 4 days before the end of a 10-day course of standard antimicrobial therapy such as vancomycin. In one study, this combination reduced the relapse rate in patients with previous relapses compared with use of vancomycin alone.93,96 However, results with this approach were not as good as published studies examining the tapering vancomycin regimen above,92,93 and there is no evidence as yet that the concomitant use of S boulardii with antibiotics alters diarrhea or prevents the appearance of C difficile toxin in the stool. Saccharomyces boulardii is not available in many countries, including the United States, but Saccharomyces cerevisiae (brewer’s yeast) is more widely available, and has been used successfully in patients with refractory CDAD.97,98 A small number of patients with refractory C difficile disease have responded to antimicrobial therapy in conjunction with an infusion of intravenous immunoglobulin at a dose of 200 to 300 mg/kg,99 or to orally administered bovine colostrum antibody harvested from cows immunized against C difficile.100-103 It is not yet accepted therapies. Similarly, reconstitution of intestinal flora through bacteriotherapy has occasionally been used in the management of individuals with refractory CDAD.105,106

CONCLUSIONS

Clostridium difficile is the most common cause of nosocomial diarrhea and is a leading cause of diarrhea in the elderly. The antibiotics most frequently associated with infection are clindamycin, ampicillin and/or amoxicillin, and cephalosporins; however, all antibiotics have been associated with CDAD, and nonessential antibiotic prescription should be avoided.107 The clinical presentation may vary from mild diarrhea to severe debilitating infection. In some cases, life-threatening complications may be accompanied by a decrease in diarrhea due to ileus. Education of hospital personnel, appropriate diagnostic testing, early treatment, and measures for preventing spread of the pathogen may avert morbidity and mortality.

Accepted for publication August 22, 2000.

This work was partially supported by grants AI01332 (Dr Ryan) and AI40725 (Dr Calderwood) from the National Institute of Allergy and Infectious Diseases, Bethesda, Md. Dr Mylonakis is the recipient of a postdoctoral research fellowship for physicians from the Howard Hughes Medical Institute, Chevy Chase, Md.

We thank Rex Neal Smith, MD, for his assistance with pathological specimens.

Corresponding author and reprints: Stephen B. Calderwood, MD, Division of Infectious Diseases, Gray-Jackson 504, Massachusetts General Hospital, 55 Fruit St, Boston, MA 02114 (e-mail: scaldwell@partners.org).

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(Reprinted) Arch Intern Med/Vol.161, Feb 26, 2001 WWW.ARCHINTERNMED.COM

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